

Enzyme Assisted Preparation of Enantiomerically Pure β -Adrenergic Blockers I. A Facile Screening Method for Suitable Biocatalysts

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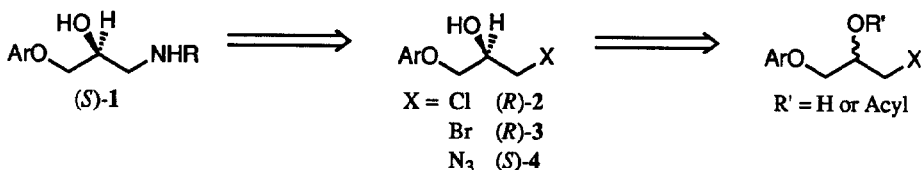
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Abstract: The enantioselective preparation of potential precursors for β -adrenergic blockers was studied by enzymatic hydrolysis of the corresponding ester derivatives. Based on the facile separation of both substrates and products by HPLC on a chiral support a convenient screening method was developed for a rapid evaluation of the best suited esterhydrolase.

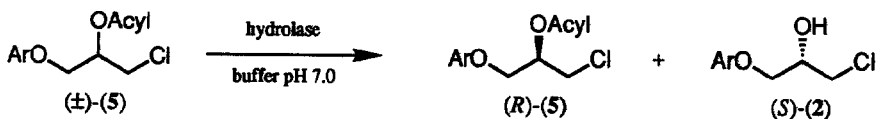
It is well established that the biological activities of β -adrenergic blockers of the general formula 1 reside largely in the (*S*)-enantiomers¹ of these molecules. Consequently, numerous attempts have been made to prepare these compounds in enantiomerically pure form, e.g. by asymmetric synthesis² or biotransformations³.

(*S*)-1 can be correlated retrosynthetically with a number of potential precursors, e.g. the corresponding α -chloro-, α -bromo- or α -azido-derivatives (Scheme 1), which in principle all could serve as starting materials for the title compounds.



Scheme 1

Esterhydrolases (Esterases, Lipases) are well known for their capability of enantiomer differentiation. In view of our previous experience⁴ in this area and based on literature data⁵, we felt that the enantioselective hydrolysis (Scheme 2) or synthesis of the corresponding esters⁶ could well provide a facile route to the desired intermediates in optically pure form.



Scheme 2

For this purpose, however the most suitable

a) biocatalyst

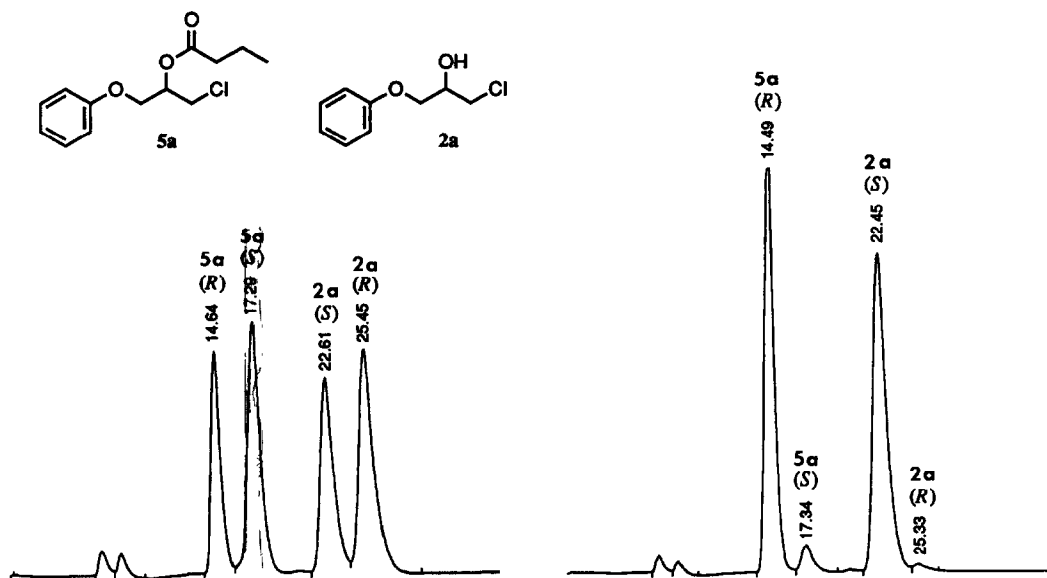
b) intermediate

first had to be found.

Target orientated screening for suitable enzymes is usually a very time consuming and tedious enterprise. We have therefore decided to develop a facile screening system for this purpose based on the enantioselective separation of the reaction products by HPLC on chiral supports. Using a commercially available column (Chiralcel OB)⁷ all products resulting from the enantioselective, enzymatic hydrolyses (Scheme 1) can be separated simultaneously. A direct determination of the achieved conversions, as well as configurations and enantiomeric purities of both substrates and products, can be achieved in one single experiment. The obtained data allow a rapid determination of the enantioselectivities⁸ displayed by the employed biocatalysts and thus a rapid evaluation of their synthetic usefulness.

lipase from *Candida sp.*

lipase from *Pseudomonas sp.*



Scheme 3

The results obtained in the hydrolyses of 2-butyloxy-1-chloro-3-phenoxypropan [(±)-5a] with a series of biocatalysts are summarized in Table 1 and exemplified in Scheme 3 for two lipases of widely different enantioselectivities.

Both from Table 1 and Scheme 3 it is obvious that the best suited biocatalysts are the lipases from *Mucor miehei* and *Pseudomonas sp.*. In view of the much higher specific activity the lipase from *Pseudomonas sp.* was chosen for further experiments.

Table 1

$(\pm)\text{-(5a)} \xrightarrow[\text{buffer pH 7.0}]{\text{enzyme}} (\text{R})\text{-(5a)} + (\text{S})\text{-(2a)}$

enzyme	t (25%) [h]	products	% e.e.	conv. ⁷	E^7
lipase from <i>Aspergillus oryzae</i>	-	$(\pm)\text{-(2a)}$	-	-	-
lipase from <i>Aspergillus niger</i>	-	$(\pm)\text{-(2a)}$	-	-	-
lipase from <i>Aspergillus sojae</i>	12	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	19.6 24.7	0.44	2
lipase from <i>Candida lipolytica</i>	74	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	70.7 90.5	0.44	42
lipase from <i>Candida sp.</i>	2	$(\text{S})\text{-(2a)}$ $(\text{R})\text{-(5a)}$	20.7 12.7	0.62	<2
lipase from <i>Chromobacterium viscosum</i>	<2	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	66.9 75.6	0.47	14
lipase from <i>Humicola lanuginosa</i>	28	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	61.0 94.0	0.39	60
lipase from <i>Geotrichium candidum</i>	12	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	48.2 60.2	0.44	6
lipase from <i>Mucor miehei</i>	53	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	44.4 98	0.31	>100
lipase from <i>Penicillium roquefortii</i>	-	$(\pm)\text{-(2a)}$	-	-	-
lipase from <i>Rhizopus delemar</i>	52	$(\text{S})\text{-(2a)}$ $(\text{R})\text{-(5a)}$	20.9 12.7	0.62	2
lipase from <i>Porcine pancreas</i> (PPL)	19	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	43.7 82	0.35	15
cholesterinesterase from <i>Pseudomonas fluorescens</i>	<1	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	24.6 27.6	0.47	2
lipase from <i>Pseudomonas sp.</i>	<2	$(\text{R})\text{-(2a)}$ $(\text{S})\text{-(5a)}$	86.2 96.3	0.47	>100
porcine liver esterase (PLE)	6	$(\text{S})\text{-(2a)}$ $(\text{R})\text{-(5a)}$	14.0 1.5	0.90	1

In view of the convenient accessibility and synthetic usefulness the α -chloro-, α -bromo- and α -azido-intermediates were chosen for the following preparative studies.⁶

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